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## Growth and Physiological Mechanism of *Phoebe chekiangensis* Seedlings in Response to Lead Stress

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### ABSTRACT

Experiments were conducted on 1-year *Phoebe chekiangensis* seedlings treated by different concentration (0, 300, 600, 900, 1200 mg/L) of Pb (NO<sub>3</sub>)<sub>2</sub>. Sixty days later, determination was implemented on seedling growth, physiological and photosynthetic parameters. The results showed that the lower concentration treated could promote the growth of the seedlings. But with the increase of concentration of lead, *P. chekiangensis* seedling height increment, ground diameter growth, whole biomass, total root surface area, root volume, total root length and root activity decreased, while root-shoot ratio present a rising trend. With the increase of concentration of Pb(NO<sub>3</sub>)<sub>2</sub> solution, the membrane permeability and MDA content of *P. chekiangensis* seedlings showed a trend of rise after the first reduce; the protein content and chlorophyll content presented a trend of decrease after the first increase; while the POD, SOD and CAT activity increased firstly but decreased afterwards; the net photosynthetic rate, stomatal conductance, intercellular CO<sub>2</sub> concentration, transpiration rate were all increase at first then decrease, which indicated that protection enzyme activity and membrane was damaged thus the growth of *P. chekiangensis* seedlings was inhibited.

### Introduction

Heavy metal pollution becomes a serious problem in the world which could be due to the anthropogenic activities such as industrial waste, fertilizers, sewage sludge application, dust from smelters and bad watering practices in the agriculture lands. However, heavy metals are not degraded by any process in ecosystems, only transfer by plants from abiotic to biotic system (Sharma et al., 2007; Mukesh et al., 2008) and

threat to plants and animals and ultimately on the health of human beings via the food chain. Lead is one of the most common heavy metals contaminants in the environment. Phytoremediation has been generally considered as a cost-effective and environmental friendly method for cleaning heavy metal polluted soils (French et al., 2006). The mechanisms and application of phytoremediation has been studied in recent years (Seregin and Kozhevnikova 2008; Yan and Tam 2011; Arya et al., 2013; Lamhamdi et al., 2013;

Kasim et al., 2014; Wińska-Krysiak et al., 2015; Zhou et al., 2017).

With the accumulation of Pb in the soil, lead could be taken up and enrichment in plants because Pb is a nonessential element for plants. When Pb has been absorbed by plants, Pb can disturb the acquisition and transport of mineral nutrition, inhibit the photosynthetic processes, accelerate free radicals and reactive oxygen species (ROS) production then causing oxidative stress in plants, reduce the enzyme activities and damage the membrane permeability, thereby impedes plants growth and development (Sharma and Dubey, 2005). The initial response of plants is the excessive generation of free radicals and ROS in plant tissues upon exposure to high levels of lead, like many other heavy metals. These ROS can lead to irreversible oxidization of lipids, proteins, chloroplastic pigments, nucleic acids, and thus affect cell viability (Foyer et al., 1994; Malecka et al., 2001; Verma and Dubey, 2003). Plants trigger both enzymatic and non-enzymatic antioxidative systems to detoxify the cells (Dalton, 1995). The antioxidant enzymes, such as superoxide dismutases (SOD), catalase (CAT), peroxidases (POD), can reduce the cellular oxidative damage and increase resistance to heavy metals (Michalak, 2006).

*Phoebe chekiangensis* C. B. Shang belongs to Lauraceae family, which was discovered by Chinese plant taxonomists Xiang Qibai in Zhejiang province (Xiang, 1974). It has been listed as rare and endangered species in china. *P. chekiangensis* is not only the good timber for premium furniture, carving, and precision mould, but also a potential ornamental plant in gardens and landscapes for its evergreen, tall shape, majestic crown, fast-growing and resisting wind. The previous studies were mainly focused on the growth affected by cultivation (Li and Xiang, 2004; Li and Xiang, 2006; Wang et al., 2013ab; Li et al., 2015; Qiu et al., 2016), population structure (Zang et al., 2015) and tissue culture (Huang, 2017). However, it was a blank on the heavy metal stress in *P. chekiangensis*. Lead is one of the ubiquitously distributed most

abundant toxic elements in the soil (Yadav, 2010). Therefore, it is necessity to study on the response of *P. chekiangensis* under the Pb stress and screen out more potential plants for phytoremediation.

The purposes of this study were aimed to understand the effects of Pb stress on growth, physiological characteristics and photosynthetic parameters in *P. chekiangensis* seedlings, and explore the possible mechanisms in the Pb stress. The results will provide information and scientific guidance on cultivation and landscape application of *P. chekiangensis*.

## Materials and Methods

### Experimental materials and preparation method

*Phoebe chekiangensis* seeds were collected from *Phoebe chekiangensis* test site in Jingmen, Hubei province in November 2014. Seeds were depositing for one month after taking out the seed coat. Sown seeds into the sowing media (3:1:1 loam: sand: sawdust, v/v/v) in March 2015. After germination, the 1-year healthy and uniform seedlings were transferred into plastic pots (8 cm deep, 8 cm in diameter) in a greenhouse with a light period from natural light and temperatures of 28/25°C (day/light). The experiment was conducted Pb stress processing after 30d.

A randomized complete block design was employed in the experiment. Five different Pb concentrations (0, 300, 600, 900, 1200mg/L) from Pb nitrate were poured into the soil for once every week lasting for 60d. The volume was 150 mL per pot every time. A total of 20 seedlings (5 per replicate) were used for each treatment. After last irrigation of Pb nitrate, cultured for 7d and then measured each growth and physiological index.

### Growth and enzyme

Seedling height and ground diameter were measured at before and 7d after Pb treatment. The seedling height increment (SHI) and ground diameter increment (GDI) were reflected by the

difference values between the first and second measurements.

Three seedlings were chosen randomly for each treatment, then carefully washed deionized water after rinsing with tap water, dried at 80°C for 48h and kept at 60°C in an oven until completely dried for biomass weigh (shoot and root).

Fresh roots were harvested for determination of root activity and root growth indicators: root surface area, the root volume, the total number of root tip and the total root length using the following procedures.

Root activity was measured by triphenyltetrazolium chloride (TTC). Took 0.25 mL 0.4% TTC solution into 10mL-volumetric flask, added a small amount of Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub> and diluted with ethyl acetate to volume, mixed sufficiently to produce red TTF.

Transferred 0.25, 0.50, 1.00, 1.50 and 2.00mL TTF solution to 5 10mL-volumetric flask separately, added ethyl acetate to volume and mixed to get 25, 50, 100, 150, 200µg TTF standard colorimetric series, with a blank as the reference, to determine OD of the solution in the spectrophotometer 485nm, then with TTC concentration as the abscissa, OD as ordinate, drew standard curve.

Immersed 0.5g plant root samples in 10mL beaker with the mixed liquid of 0.4% TTC and 66mmol/L phosphate buffer solution (pH=7.0), kept at 37°C for 1h, then added 1mol/L of sulfuric acid 2mL to terminate reaction. Removed the root, carefully wiped it and grinded with 3-4mL ethyl acetate and a small amount of quartz sand in a mortar to extract TTF, used a small amount of ethyl acetate to wash the residue for 2-3 times, then poured into the test tube, finally added ethyl acetate to 10mL, used a spectrophotometer for 485 nm colorimetry, taking the blank test (with sulfuric acid, then added the root samples) as reference, readout the OD. According to the standard curve, the reducing amount of TTC, we can get the reducing strength of TTC.

Reducing Strength of TTC= Reduction Amount of TTC (µg) / [Weight of Root Sample (g) × Time (h)]

Chosen 3 root samples from each treatment for root growth indicators measurement by Epson expression 10000xl 1.0 root system scanner and analysis by WinRHIZO Pro 2005b.

Fresh leaves were harvested for further measurement of membrane permeability, Chlorophyll content, soluble protein, MDA, POD, SOD and CAT using the following procedures.

Membrane permeability determination was according to Niu's method. Took 1.5g tested plant leaf treated by different concentration, washed it carefully by distilled water and cut it as square, put into a 100mL triangle flask, added 40mL distilled water, shook and kept the leaves were all immersed in distilled water for 24h at room temperature on a shaker, measured the electrical conductance value E1. Then put into boiling water for 30 min, kept 24h at room temperature and measured the electric conductivity E2. Used E1/E2 × 100% to express the relative membrane permeability (%).

The determination procedure of chlorophyll content as following: 0.2g of fresh leaf blades were cut into small pieces and placed into 10mL centrifuge tube with plug, filled with 95% ethyl alcohol to scale, kept in darkness for 72h, until the leaf pieces were completely white.

The liquid supernatant was measured the chlorophyll content. The chlorophyll in the solution was determined with a spectrophotometer at 665 and 649 nm. Chlorophyll content was calculated according to the following formulae:

$$C_a = 13.95A_{665} - 6.88A_{649}$$

$$C_b = 24.96A_{649} - 7.32A_{665}$$

$$C_{a+b} = (C_a + C_b) \times V \times N / W / 1000$$

Where C<sub>a</sub> is the chlorophyll a content, C<sub>b</sub> is the chlorophyll b content, C<sub>a+b</sub> is the total chlorophyll content, and A is the absorbance at the specified wavelength.

To prepare enzyme extraction, about 0.2g of leaf samples were homogenized with 1.6mL of 150mM buffer solution (containing 0.7 of  $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$  and 1.64%  $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$ , pH 7.8) subjected to grinding with an ice-cooled mortar and pestle and finally centrifuged at 12000rpm for 20 min at 4°C. The supernatant was collected for the determination of antioxidative enzyme activities, as well as the content of soluble protein and MDA. SOD activity was assayed following the procedure described by Dhindsa et al., (1981). CAT and POD activity was determined according to Chance and Maehly (1955). The MDA content was established by the method of Heath and Packer (1968). The soluble protein content was estimated as described by Bradford (1976).

### Determination of photosynthetic parameters

A Li-6400 portable photosynthesis meter (Li-Cor Inc. USA) was used to measure the following parameters for *Phoebe chekiangensis* seedlings from each treatment: photosynthetic rate (Pn), transpiration rate (Tr), stomatal conductance (Gs), and mesophyll intercellular  $\text{CO}_2$  (Ci). The built-in light source and  $\text{CO}_2$ -supplying system were utilized to provide light energy and  $\text{CO}_2$  fixed in photosynthesis.

Photosynthetic active radiation was set at  $1200 \mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ . In addition,  $450 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$  was set as the external  $\text{CO}_2$  level, at which the photosynthesis measurements were made, according to the seedling-growing conditions. During the experiment, intact mature function leaf blades were chosen from the upper part of the shoots. Data from 8 replicates were used to determine all parameters.

### Statistical analysis

Data were analyzed by one-way ANOVAs with the Duncan's multiple range tests to separate means, using the program SAS 10.0 (SAS Institute, Inc., Cary, NC). The correlation analysis was conducted using SPSS 18.0. Different letters in graph or tables indicate significant differences at  $p < 0.05$ .

## Results and Discussion

### Growth of *Phoebe chekiangensis* seedlings under different Pb concentrations

After the Pb stress treated, the seedling height increment, ground diameter increment, biomass and root-shoot ratio of *Phoebe chekiangensis* seedlings were significant influenced by the Pb stress (Table 1). The seedling height increment, ground diameter increment and biomass increased firstly and then decreases as Pb concentration increased while root-shoot ratio showed a converse trend.

At a concentration of 300mg/L, SHI, GDI and biomass reached their highest values, which were 33.33, 21.67, and 10.10% higher than the treatment without Pb stress (the control), respectively. At 600mg/L of Pb, SHI and GDI were also significant higher than the control, were 13.69 and 10.00%, respectively. Under T3 and T4 treatments, the decreases were showed on the SHI, GDI and biomass, which were 20.23, 16.67, 18.68% and 27.98, 26.67, 27.02% lower than the control, respectively. There were no significant affected on the root-shoot ratio compared with the control. The root-shoot ratios under all treatments expect 300mg/L of Pb were higher than the control.

### Root growth of *Phoebe chekiangensis* seedlings under different Pb concentrations

As the increasing of Pb concentration, the total root tips, root surface area, root volume, total root length and root activity of *Phoebe chekiangensis* seedlings were climbed up and then declined (Table 2). At Pb concentration of 300mg/L, all of the indicators reached their highest values, which were 67.49, 13.47, 41.72, 19.61, and 15.24% higher than the control, respectively. Among them, the Pb stress had a significantly increased on the total root tips, total root length and root activity. Under 600mg/L of Pb, root activity was significantly higher than the control, while the other indicators were lower than the control. Compared with CK, Pb was significantly deceased total root length and root activity.

### Physiological characteristics of *Phoebe chekiangensis* seedlings under different Pb concentrations

As the Pb concentration increased, the changes of physiological characteristics of *Phoebe chekiangensis* seedlings were shown in Fig. 1. The membrane permeability and MDA content decreased firstly then increased with an increase in Pb concentration. In addition, the differences among different treatments were significant except that of MDA content between the control and 600mg/L. The lowest values of membrane permeability and MDA content were 64% and  $0.016\mu\text{mol}\cdot\text{g}^{-1}\cdot\text{Fw}$  under 300mg/L concentration treated, which were 5.88% and 26.39% significantly lower than that in the control. As well as the membrane permeability was 1.47% significantly lower than the control at a concentration of 600mg/L. At 900mg/L and 1200mg/L of Pb concentration, the membrane permeability and MDA content were significantly higher than that of the control by 2.94%, 4.41% and 21.24%, 46.65%, respectively.

The soluble protein and Chlorophyll content in the treated plants leaves were shown as an increased firstly then decreased trend. At the Pb concentration from 300 to 900 mg/L, the soluble protein and chlorophyll content were higher than that in the control. The highest values were reached at 600mg/L Pb, and were 1.4184 and 3.1694mg/g which were 87.50 and 41.12% higher than that of the control, respectively. Beside for the soluble protein content at 600mg/L concentration, all treatments had no significant difference of control. But at 300 and 600 mg/L concentration, the soluble protein and chlorophyll content were significantly higher than that at 1200mg/L.

The effect of different Pb concentration on the antioxidant enzyme activities of *Phoebe chekiangensis* seedlings was shown in Fig. 1. As an increase of Pb concentration, all of the antioxidant enzyme activities were increased firstly then decreased. At 300mg/L and 600mg/L concentration, the SOD, CAT, and POD activities

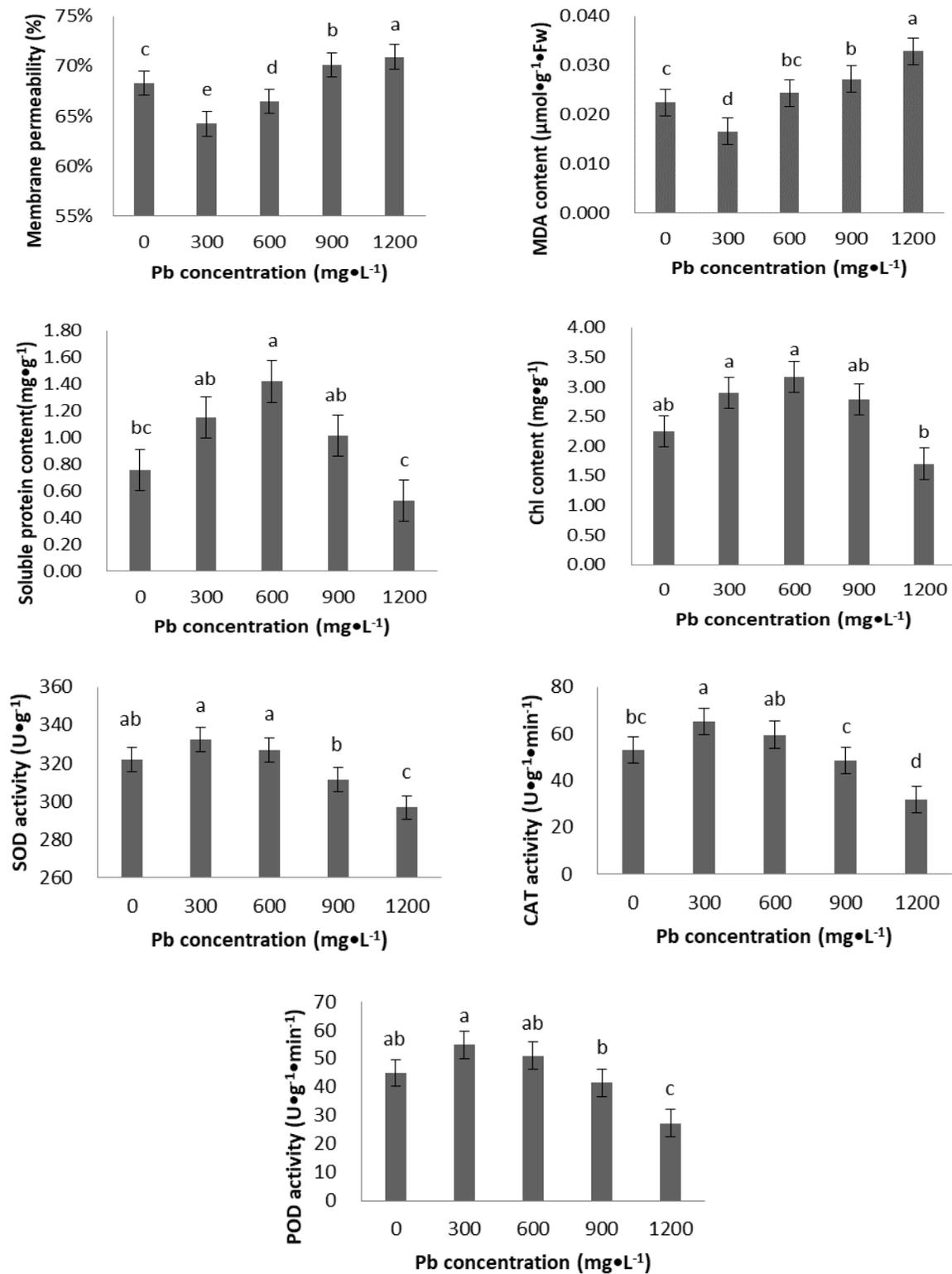
were higher than that of the control. And the highest values were 332U/g, 65.0862U/(g·min), and 54.7725U/(g·min) at 300mg/L Pb concentration, respectively, and higher than that of the control by 3.20, 22.34, and 21.84%, respectively. There were significant differences between CAT activity and the control at 300 mg/L Pb concentration, while all of the antioxidant enzyme activities were significantly lower than that of the control under a concentration of 1200mg/L treated.

### Photosynthetic parameters of *Phoebe chekiangensis* seedlings under different Pb concentrations

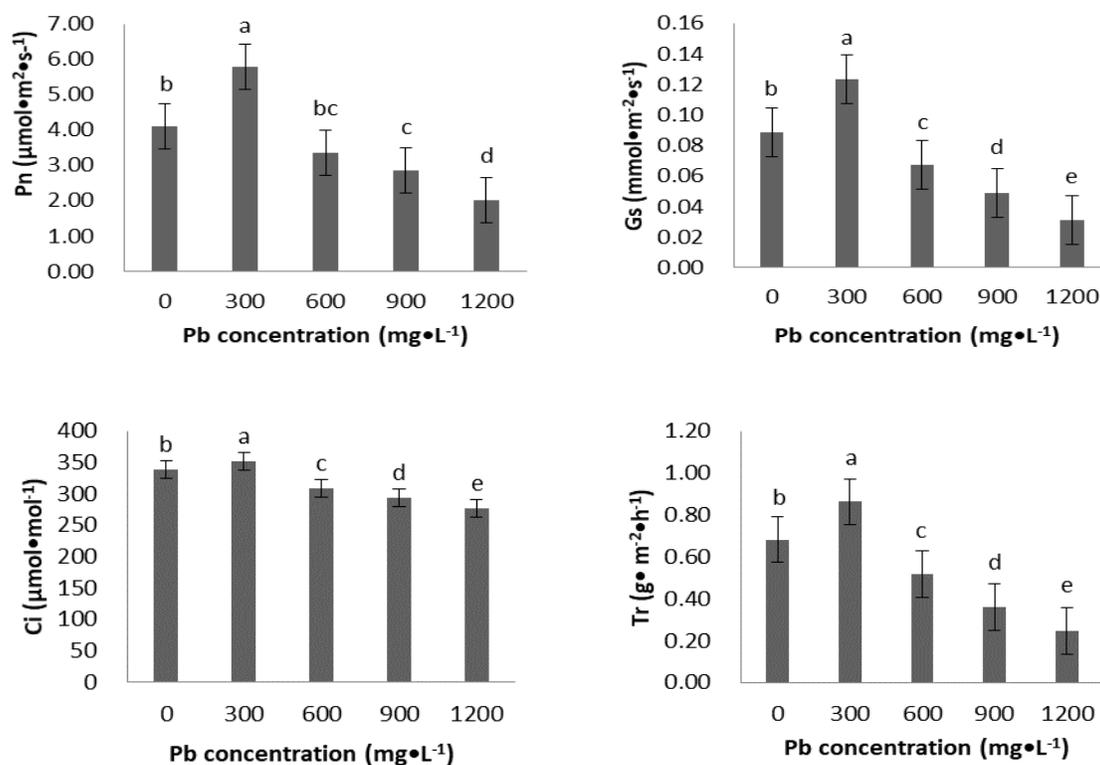
In this experiment, Pb had significant influence on photosynthetic parameters. Most of the differences between treatments were also significant (Fig. 2). All of the photosynthetic parameters showed an upward firstly then downward trend as Pb concentration increased. The Pn, Gs, Ci, and Tr reached their highest values in seedlings treated with 300mg/L and were 41.38%, 39.01%, 3.94%, and 26.30% higher than that in the control. As the Pb concentration increased continuously, the Pn, Gs, Ci, and Tr were generally reduced. Each treatment value was significantly lower than the control value, except the Pn under 600mg/L treatment.

### Growth response of *Phoebe chekiangensis* seedlings under Pb stress

Plant sensitivity and response to heavy metal Pb depend upon the plant genetic and physiological characteristics (Singh et al., 1997). Some plant species which have higher Pb accumulation have no bad influence on the growth under Pb stress, even can be promoted (Wierzbicka, 1995). Plant growth is the most observable reflection of the plant toxicity by heavy metals stress. In our present study, the growth of *P. chekiangensis* seedlings as indicated by SHI, GDI and biomass was promoted by the lower Pb concentration while inhibited by the higher Pb concentration, which was in agreement with the results by Lin and co-workers (Lin et al., 2017).



**Fig. 1:** Effects of physiological characteristics of *Phoebe chekiangensis* seedlings under different concentrations of Pb (NO<sub>3</sub>)<sub>2</sub> stress. The same alphabets between treatments did not differ significantly at 5% level by Duncan's multiple range test.



**Fig. 2:** Effects of photosynthesis parameters of *Phoebe chekiangensis* seedlings under different concentrations of Pb (NO<sub>3</sub>)<sub>2</sub> stress. The same alphabets between treatments did not differ significantly at 5% level by Duncan’s multiple range test.

**Table 1.** Effects of the growth of *Phoebe chekiangensis* seedlings under different concentrations of Pb(NO<sub>3</sub>)<sub>2</sub> stress.

No.	Pb concentration (mg/L)	Seedling height increment (cm)	Ground diameter increment (cm)	Biomass (g)	Root-shoot ratio
T0	0	1.68 c	0.060 c	4.57 b	1.31 ab
T1	300	2.24 a	0.073 a	4.97 a	1.29 b
T2	600	1.91 b	0.066 b	4.71 ab	1.33 ab
T3	900	1.34 d	0.050 d	3.72 c	1.35 ab
T4	1200	1.21 e	0.044 e	3.34 d	1.39 a

The same alphabets between treatments did not differ significantly at 5% level by Duncan’s multiple range test.

**Table 2.** Effects of the root growth of *Phoebe chekiangensis* seedlings under different concentrations of Pb (NO<sub>3</sub>)<sub>2</sub> stress.

No.	Pb concentration (mg/L)	Total root tips	Root surface area (cm <sup>2</sup> )	Root volume (cm <sup>3</sup> )	Total root length (cm)	Root activity
T0	0	808.67 b	18.19 ab	1.63 ab	253.14 b	327.79 c
T1	300	1354.67 a	20.64 a	2.31 a	302.78 a	377.64 a
T2	600	725.33 b	17.32 ab	1.35 ab	227.84 bc	344.64 b
T3	900	618.33 b	16.33 b	1.20 b	213.42 c	301.16 d
T4	1200	490.67 b	15.73 b	0.96 b	198.71 c	139.76 e

The same alphabets between treatments did not differ significantly at 5% level by Duncan’s multiple range test.

On the contrary, the root-shoot ratio had a downward firstly then upward trend. When under the Pb stress, plants distribute the limited nutrition to different functional organs to help adaptation or against the stress. In our study, in order to resist the Pb stress, *P. chekiangensis* seedlings transferred the nutrition to the roots to facilitate the root growth for more nutrients. As a result, the root biomass rise up and the root-shoot ratio gradually increased.

### **Root growth response of *Phoebe chekiangensis* seedlings under Pb stress**

Roots are the most important organs for absorption of water and nutrients in plants. Heavy metals inhibit the root cell mitotic, affect root system development, and influence the roots to absorb water and nutrients. In our experiment, the root growth indicators were shown an increase firstly then decrease trend. These results agreed with the reports of *Machilus pauhoi* Kanehira (Zheng et al., 2015). The roots are the earliest tissues directly in contact with soil surrounding the Pb and also the location where the heavy metals have been taken up into the plants. Therefore, plant root plays an important role in tolerance mechanism.

In the previous research, there are two tolerance mechanisms related with plant roots. One is plant root cell wall which can bind the metals with proteins present in the epidermal cells of cell wall and ultimately reduce translocation and accumulation of heavy metals in upper parts (Shameem et al., 2016). The other is root exudates which result in chelation of heavy metal ions to detoxify (Ma et al., 2001). In *Arabidopsis thaliana* seedlings, root growth was inhibited while cytoplasm and major organelles were protected from Pb toxicity, and the heavy metal Pb was accumulated in the root cell wall (Ing et al., 2011). However, in our study, the root growth has been promoted under the lower Pb concentration. How an action of Pb and root cell wall might impact on root growth of *P. chekiangensis* is not clear. Therefore, the further research of Pb tolerance mechanism in *P. chekiangensis* should focus on the root cell wall and root secretions.

### **Physiological response of *Phoebe chekiangensis* seedlings under Pb stress**

Cell membrane can control and adjust the transport and exchange of intracellular substances by its selectively permeability. The permeability value reflects the amount of soluble substance leakage within the cell membrane, so is the evaluation index of the reaction of plants to environmental damage. Results from our study showed that the membrane permeability decreased firstly then increased. It revealed that the seedlings detoxified the Pb stress by decreasing the membrane permeability at lower Pb concentration. With the Pb concentration continuing increased, the structure and function of plant cell membrane was harmed by the Pb stress, membrane stability decreased, passive leakage of ion cells and macromolecules. MDA, which is the final decomposition product of the lipid peroxidation due to oxidative stress, is also an indicator of membrane damage degree. In our study, MDA content showed a similar trend with membrane permeability.

In contrast to membrane permeability and MDA content, soluble protein and chlorophyll contents in leaf tissues increased firstly then reduced. Protein is the main component of cell and organelle, enzyme and membrane. Under the lower concentration of Pb, plant growth increased with a accompanying of a substantial increase of the cell, membrane and enzyme in plant, resulted in an increase of soluble protein content.

The growth of *P. chekiangensis* seedlings was inhibited by the Pb stress at the higher concentration, so the content of soluble protein gradually reduced. Chlorophyll plays a very important role in the plant photosynthesis. At the higher concentration of Pb, the Pb ion inhibited the generation of enzymes which take part in the synthesis of chlorophyll, induced excessive ROS and free radicals which affected the stability of chlorophyll structure, and competed with Mg ion to synthesis of chlorophyll. A lot of researches have reported the plant leaves chlorosis problem as a result of heavy metal stress (Shameem et al., 2016).

Normally, plants evolved complex antioxidant defense system to repair the damage initiated by ROS. SOD can catalyze the dismutation of  $O_2^{2-}$  into  $H_2O_2$  and  $O_2$ . The  $H_2O_2$  was then resolved by CAT and POD into  $H_2O$  and  $O_2$ . In present study, the enhancement of activities of antioxidant enzymes (SOD, CAT, POD) at the lower concentration of Pb, was an stress response which could keep the steady state level of ROS. Elevated concentration of Pb continuously, excessive ROS disturbed the homeostasis of ROS and antioxidant enzymes in plants. The activities of SOD, CAT, and POD declined and the plants were damaged by the Pb stress.

### Photosynthetic response of *Phoebe chekiangensis* seedlings under Pb stress

Stomatal and nonstomatal inhibitors are generally thought to affect the plant photosynthesis during environmental stress (Lin et al., 2017). Results from our study, the Pn, Gs, Ci, and Tr showed an increasing then decreasing trend, indicated that *P. chekiangensis* seedlings enhanced photosynthesis and produced more nutrients for plant growth under lower Pb concentration while were toxic with Pb concentration continuously increased which were in consistent with the results of other indicators in our study. At higher Pb concentration condition, Pb stress impeded the synthesis of chlorophyll, degraded chlorophyll, and altered the enzyme activities which participated in the photosynthesis and chlorophyll synthesis in cells. As a consequence, the photosynthesis of seedlings reduced, and then plant grew slowly.

In conclusions, the effects of Pb on *P. chekiangensis* seedlings were reflected at the growth, physiological, enzyme, and photosynthetic levels. Under lower concentration of Pb, all growth increased, membrane permeability and MDA content decreased, soluble protein and chlorophyll contents raised, antioxidant enzyme activities enhanced, and photosynthesis was promoted. The growth and physiological characteristics of *P. chekiangensis* seedlings were inhibited when exposed to higher concentration of Pb. Based on

these results, we suspected that *P. chekiangensis* was a Pb tolerant plant which might have an active and efficient mechanism of protecting plant against Pb stress. *P. chekiangensis* seedlings are promising for phytoremediation of Pb-polluted soils.

### Conflict of interest statement

Authors declare that they have no conflict of interest.

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